Product description

NS0 is a non-Ig secreting, non-light chain-synthesizing subclone of NS-1 (P3/NS1/1-Ag4-1). The cells are resistant to 10µM azaguanine, cells die in the presence of HAT medium. NS0 is widely used as a fusion partner to generate antibody secreting hybridoma cell lines and is also used as a model B-cell line.

The NS0 cell line is the property of MRC. CancerTools.org is unable to supply this cell line or derivatives of this cell line to commercial entities unless a license has been first secured from MRC, negotiated by LifeArc (formerly known as MRC Technology); requests for commercial use of this cell line will be forwarded to LifeArc.

Name: NS0 cell line

Parental cell: NS1 (P3/NS1/1-Ag4-1)

Organism: Mouse

Disease: Murine myeloma

Growth properties: Suspension

CRISPR Edited: No

Contributor(s)

Institute: Medical Research Council

Properties

Product format: Frozen

Unpacking and storage:

1. Check all containers for leakage or breakage.

2. Remove the frozen cells from the dry ice packaging and immediately place the cells at a temperature below -130°C, preferably in liquid nitrogen vapor, until ready for use.

Recommended medium: RPMI 1640 + 2mM Glutamine + 10% Fetal Bovine Serum (FBS)

Subculture: Maintain cultures between 3-9x100,000 cells/mL. Cells can adhere loosely to the culture vessel surface; remove by tapping against the flask. Cultures that are more than 2 days old may have to be scraped off.

Culture conditions: 37.0°C ± 1.0°C incubator with 5.0% ± 1.0% CO_2

Handling instructions

1. Please ensure that vials are frozen when received, and store at **<-130 °C long term**. When removing frozen cells from storage, it is important to minimize exposure to room temperature (15 - 25°C). If not

proceeding directly to thawing, place the cells on dry ice or in a liquid nitrogen container.
2. **Do not thaw at room temperature.** To thaw, swirl the vial quickly in a 37 °C water bath with O-ring and cap above the water to avoid contamination. Remove from the water bath with a small ice pellet remaining (this should not take more than 2 minutes) and wipe the exterior with 70% ethanol or isopropanol before transferring to a biosafety cabinet. Further steps should be conducted under aseptic conditions.
3. We strongly recommend that the volume of cell suspension is measured at this point, and a 20 uL aliquot be removed for a **viable cell count** using trypan blue or similar dye. This ensures that provided cells are viable, and the cell count can be used to determine volume of growth medium to be added to the cell suspension.
4. Transfer the remaining cell suspension to a 50 mL conical tube using a pipette.
5. Rinse the vial with 1 mL of medium and add it dropwise to the cells, while gently swirling the 50 mL tube.
6. Wash by adding 15 - 20 mL of medium **dropwise**, while gently swirling the tube.
7. Centrifuge the cell suspension at **250 x g for 5 minutes** at room temperature.
8. Carefully remove the supernatant with a pipette, leaving a small amount of medium to ensure the cell pellet is not disturbed. Resuspend the cell pellet by gently flicking the tube.
9. Gently add required volume of culture medium and transfer to a suitable cell culture flask.
10. Examine the cultures after 24 hours and split as required.

References

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Material Citation

If use of this material results in a scientific publication, please cite the material in the following manner: NS0 cell line (CancerTools.org #153797).